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What do we need to know about RAVs clinically ?

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Background

- Resistance associated variants (RAVs) or resistance associated polymorphisms (RAPs) with reduced drug susceptibility are observed in patients with chronic HCV
 - Due to the high replication rate of HCV (10^{12} / day)
 - Error-prone RNA-dependent RNA polymerase (10^{-4} to 10^{-5} per copied nucleotide)
- All possible single and double mutants are predicted to be generated multiple times each day
- Not all RAVs are fit enough to persist (but potentially under selection pressure of a respective DAA). Most NS3A RAVs disappear after EoT (HCV1b > HCV1a), while many NS5A RAVs persist
- The level of resistance *in vivo* may differ between a RAV in previously untreated patient and the same RAV selected by a DAA (= TEV) (e.g. due to compensatory mutations)

How to define a RAV ?

- RAV per specific drug or drug class
(e.g. resistance against elbasvir or any NS5A inhibitor)
- Genotypic backbone *in vitro* and *in vivo*
(e.g. genotype 1a or 1b replicons, other genotypes)

Group	GT	Position								
		K24	M28 (L28) [†]	Q30 (R30) [*]	L31	P32	S38	H58 (P58) [‡]	A92	Y93
EBR RAVs	1a	- [¶]	A, G, T	D, E, H, G K, L, R	F, M, V	- [¶]	- [¶]	D	- [¶]	C, H, N, S
	1b	- [¶]	Any	Any	Any	L	- [¶]	D	K	Any
NS5A Class RAVs	1a or 1b	Any	Any	Any	Any	Any	Any	Any	Any	Any

[†]Wild type amino acid at position 28 is M in GT1a and L in GT1b

^{*}Wild-type amino acid at position 30 is Q in GT1a and R in GT1b.

[‡]Wild type amino acid at position 58 is H in GT1a and P in GT1b

[¶]No EBR RAVs identified at this position

Note that EBR RAVs are a subset of the NS5A class RAVs

How to define a RAV ?

- RAV per specific drug or drug class
(e.g. resistance against ledipasvir or any NS5A inhibitor)
- Genotypic backbone *in vitro* and *in vivo*
(e.g. genotype 1a or 1b replicons, other genotypes)
- Level of reduced susceptibility *in vitro*
(e.g. any aa change vs >2.5-fold vs >100-fold)

NS5A RAVs - Overview

	GT 1a				GT1b	
	M28T	Q30R	L31M/V	Y93H/N	L31V	Y93H/N
Ledipasvir	>20x	>100x	>100x	>1,000x		>1,000x/-
Ombitasvir	>1,000x	>100x	<3x	>10,000x	<10x	>50x
			>100x			
Daclatasvir	>100x	>1,000x	>100x >1000x	>1,000x >10,000x	>20x	>20x
Elbasvir	>10x	>10x	>10x	>100x	<10x	>10x/-
			>100x			
Velpatasvir	<10x	<3x	>20x	>100x >1,000x		<3x/-
Odalasvir	>20x	<10x	<3x	>1,000x	<3x	<3x
						<10x
ABT-530	<3x	<3x	<3x	<10x	<3x	<3x
MK-8408	<10x	<10x	<10x	<10x	<10x	<10x

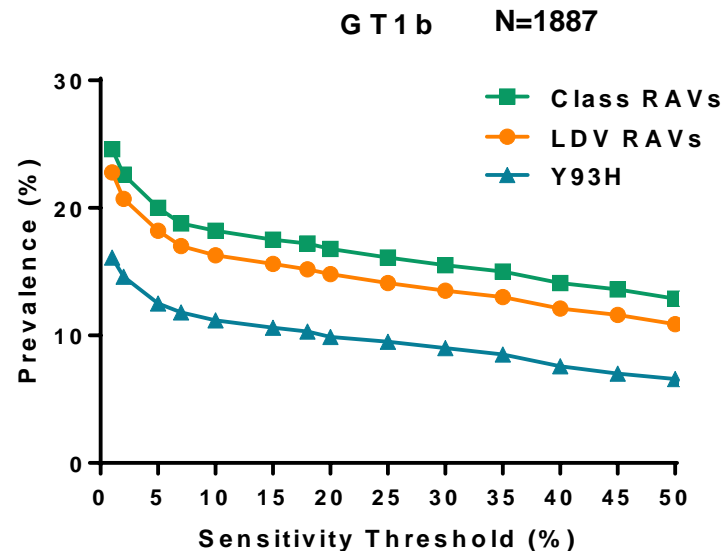
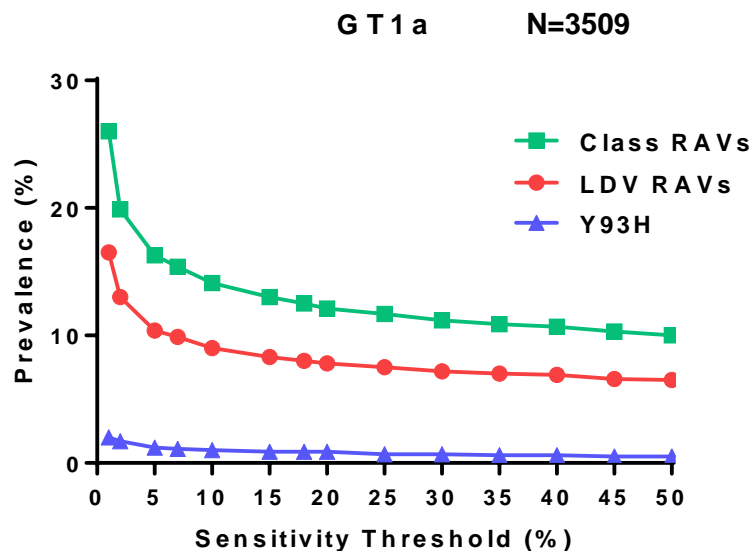
EC₅₀ fold-change compared to WT replicon

Cheng et al., EASL 2012; Wong et al., AAC 2013; Krishnan et al., AAC 2015; Fridell et al., Hepatology 2011; Liu et al., AAC 2015; Cheng et al., EASL 2013; Patel et al., EASL 2015; Ng et al., CROI 2014; Asante-Appiah et al., AASLD 2014

How to define a RAV ?

- RAV per specific drug or drug class
(e.g. resistance against ledipasvir or any NS5A inhibitor)
- Genotypic backbone *in vitro* and *in vivo*
(e.g. genotype 1a or 1b replicons, other genotypes)
- Level of reduced susceptibility *in vitro*
(e.g. any aa change vs >2.5-fold vs >100-fold)
- Methodology of RAV detection (sensitivity threshold)
(e.g. deep sequencing with a cut-off at 1% or populations sequencing with a cut-off at 15-25%)

Methodology of RAV detection



GT1a Position	LDV RAVs	NS5A Class RAVs
24	G/N/R	G/N/R
26		E
28	A/G/T	A/G/T/V
30	E/G/H/L/K/R/T	C/E/G/H/I/L/K/R/S/T/Y
31	I/F/M/V	I/F/M/V
32	L	L
38	F	F
58	D	D/L
92	K/T	K/T
93	C/F/H/N/S	C/F/H/L/N/R/S/T/W

GT1b Position	LDV RAVs	NS5A Class RAVs
28		M
31	I/F/M/V	I/F/M/V
32	L	L
58	D	D
92	K	K
93	C/H/N/S	C/H/N/S

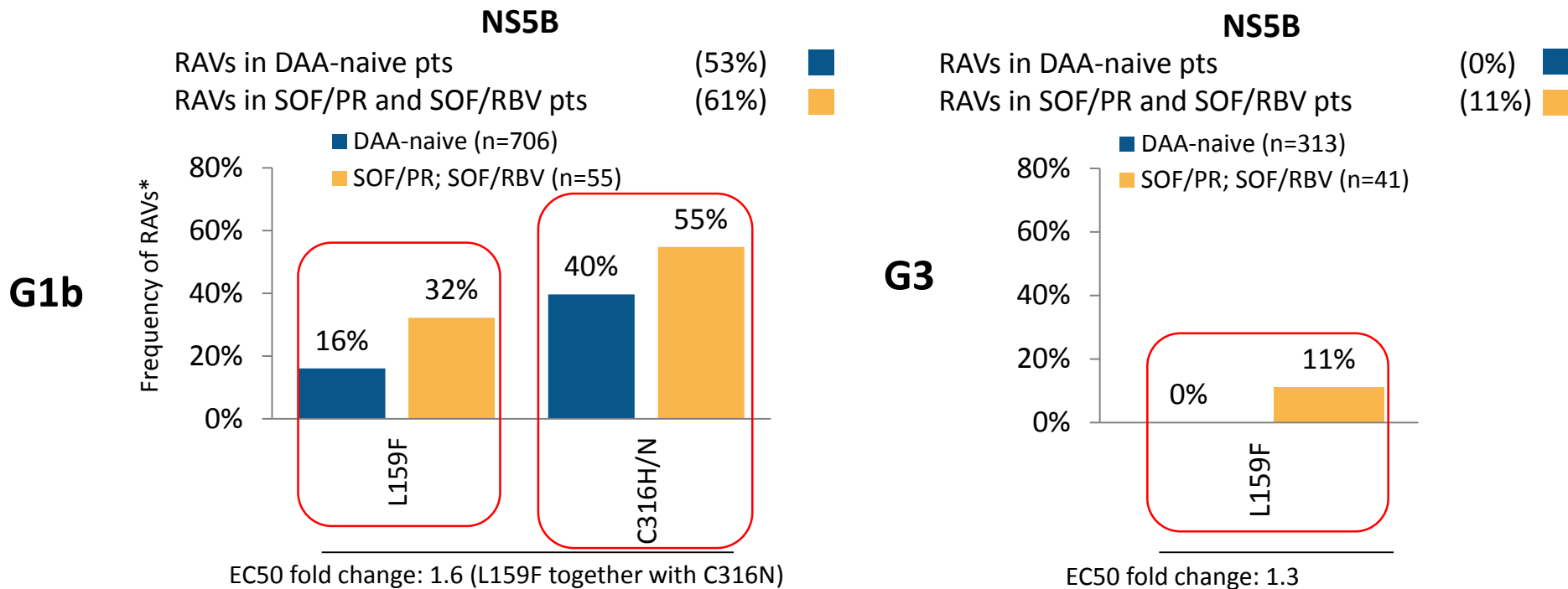
Date on file, Gilead
 Hongmei Mo, personal communication

Frequency and characteristics of RAVs in treatment-naïve and DAA-experienced patients - European RAVs database:

- Serum samples of 3549 European HCV-infected patients
- Population-based sequencing for NS3, NS5A, and NS5B
 - Considered relevant if associated with treatment failure or shown to confer >2-fold changed drug susceptibility in comparison to reference strain

RAVs in SOF/PR and SOF/RBV failures

- No RAVs detected in G1a or G2

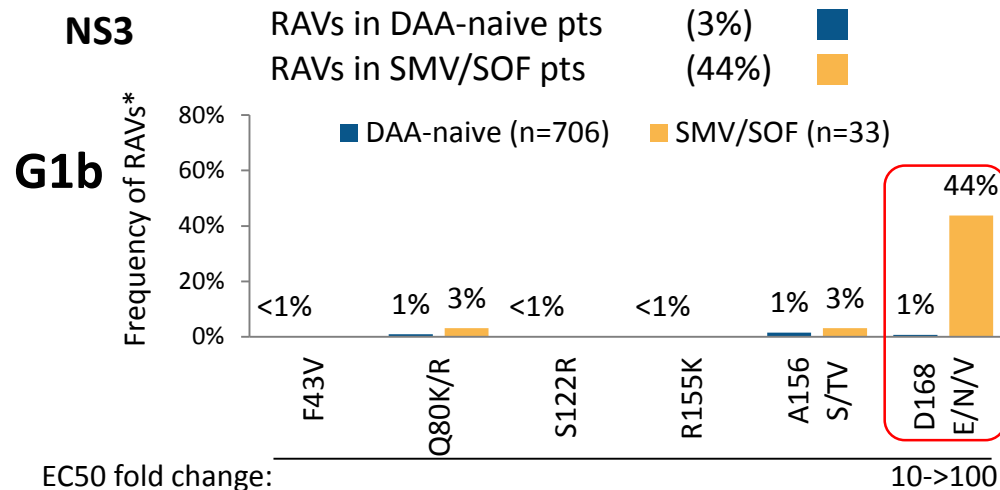
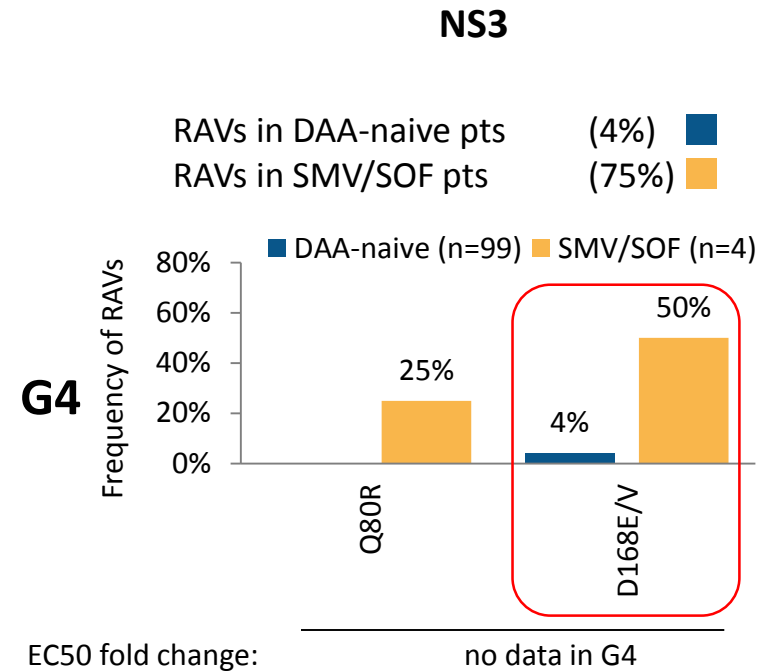
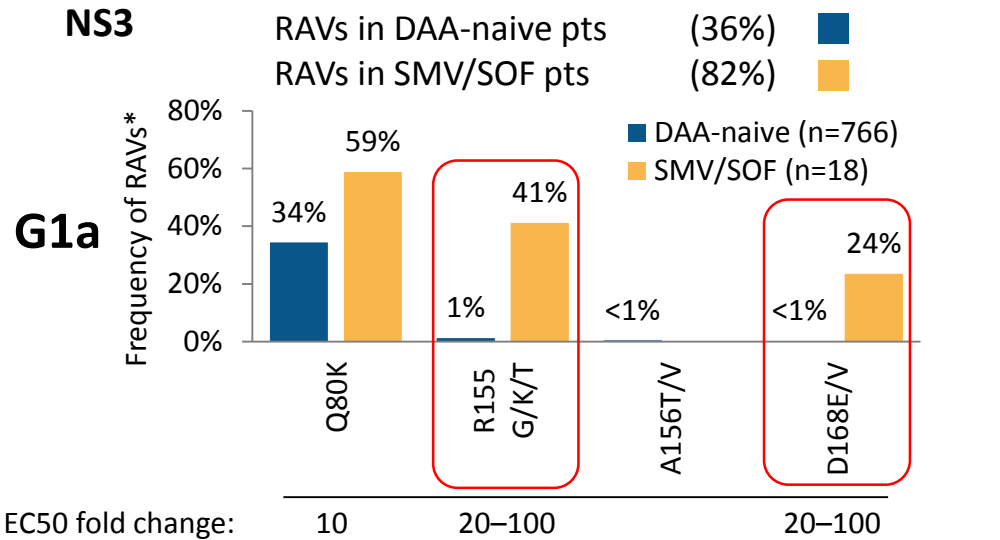


Dietz J, et al. EASL 2016, Barcelona. #PS007

*Double variants were calculated as single RAVs. Thus, sum of single RAVs frequencies is not identical with rate of patients with RAVs.

Frequency and characteristics of RAVs in treatment-naïve and DAA-experienced patients - European RAVs database:

RAVs in SMV/SOF failures



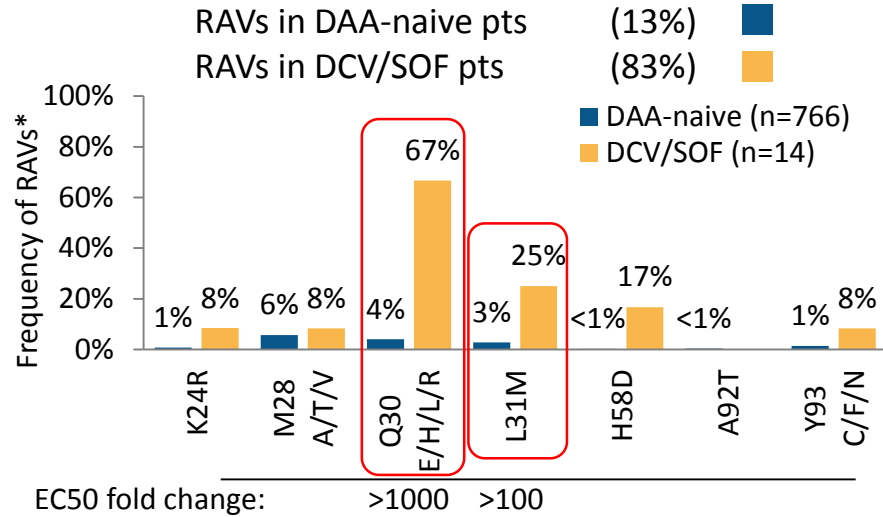
Dietz J, et al. EASL 2016, Barcelona. #PS007

*Double variants were calculated as single RAVs. Thus, sum of single RAVs frequencies is not identical with rate of patients with RAVs.

Frequency and characteristics of RAVs in treatment-naïve and DAA-experienced patients - European RAVs database:

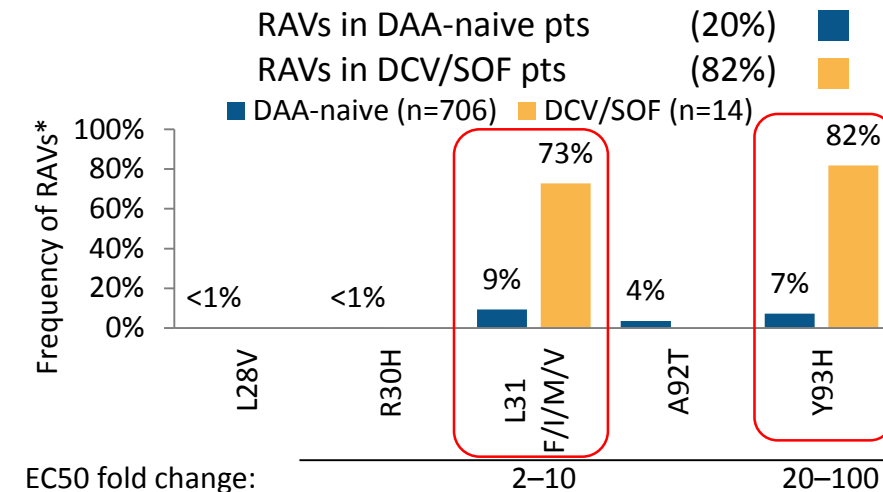
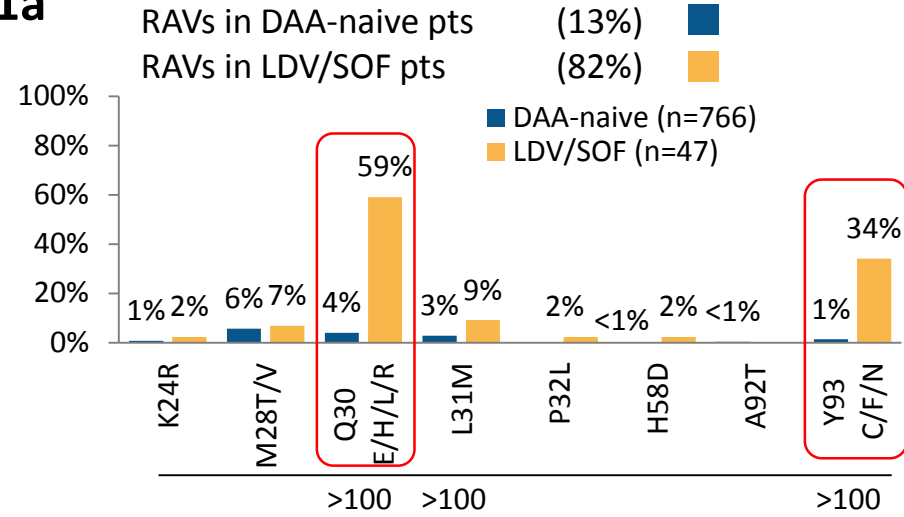
RAVs in DCV/SOF versus LDV/SOF failures (G1)

NS5A – DCV/SOF

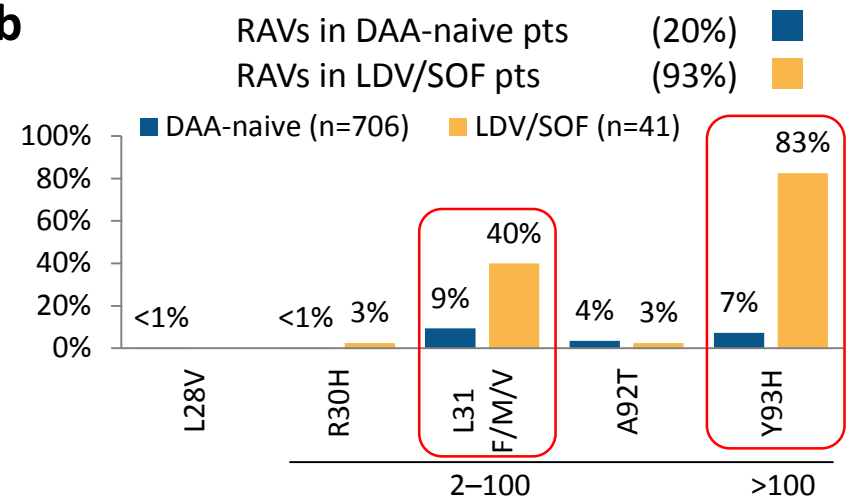


NS5A – LDV/SOF

G1a



G1b

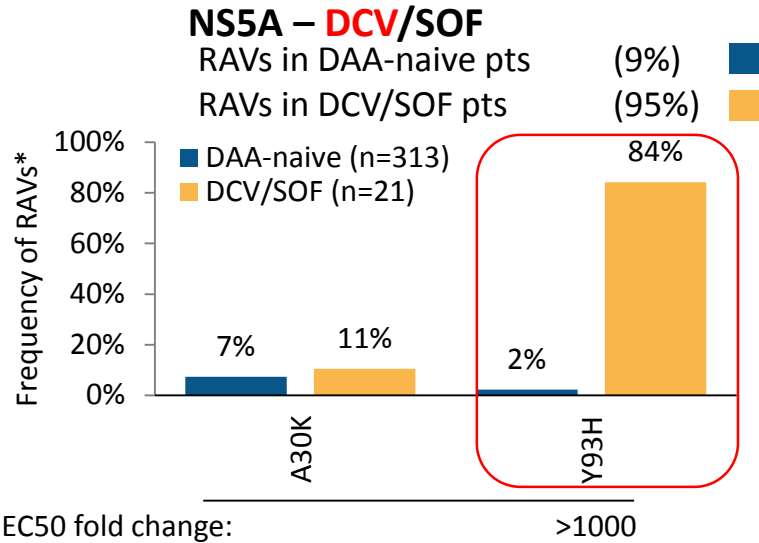


Dietz J, et al. EASL 2016, Barcelona. #PS007

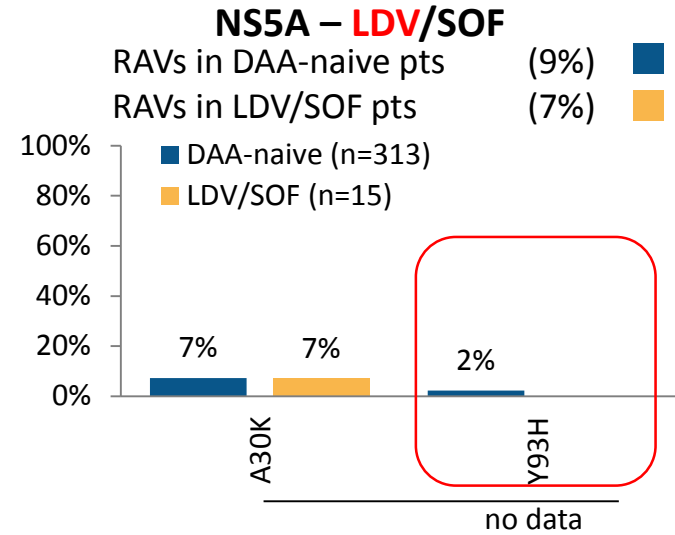
*Double variants were calculated as single RAVs. Thus, sum of single RAVs frequencies is not identical with rate of patients with RAVs.

Frequency and characteristics of RAVs in treatment-naïve and DAA-experienced patients - European RAVs database:

RAVs in DCV/SOF versus LDV/SOF failures (G3 & 4)

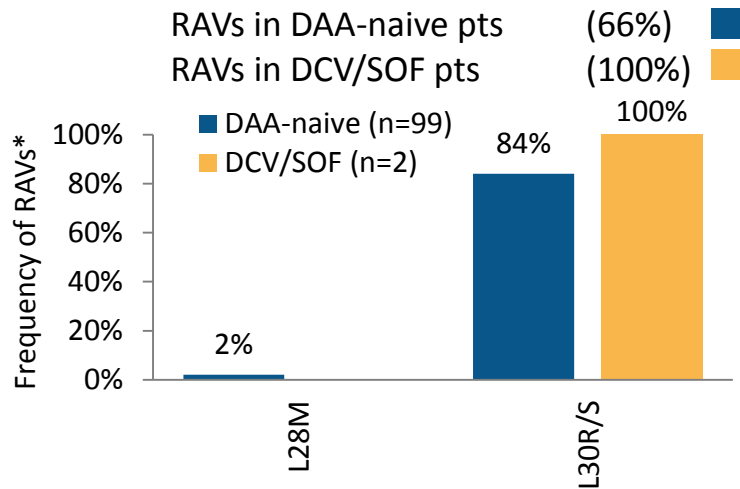


G3

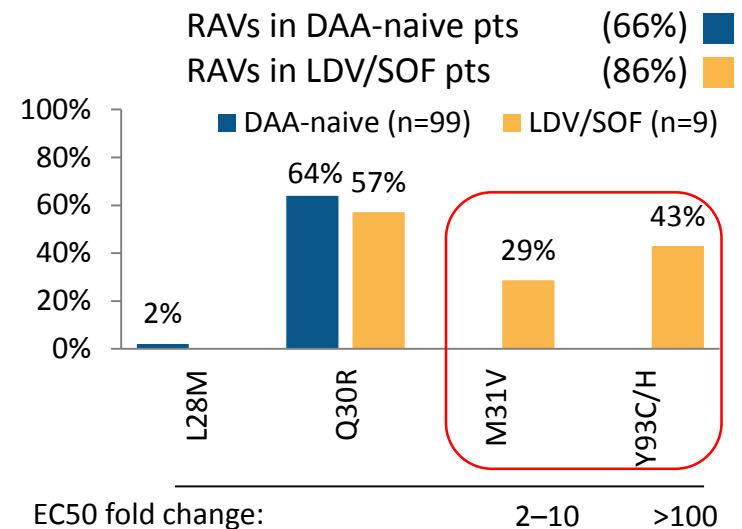


EC50 fold change:

>1000

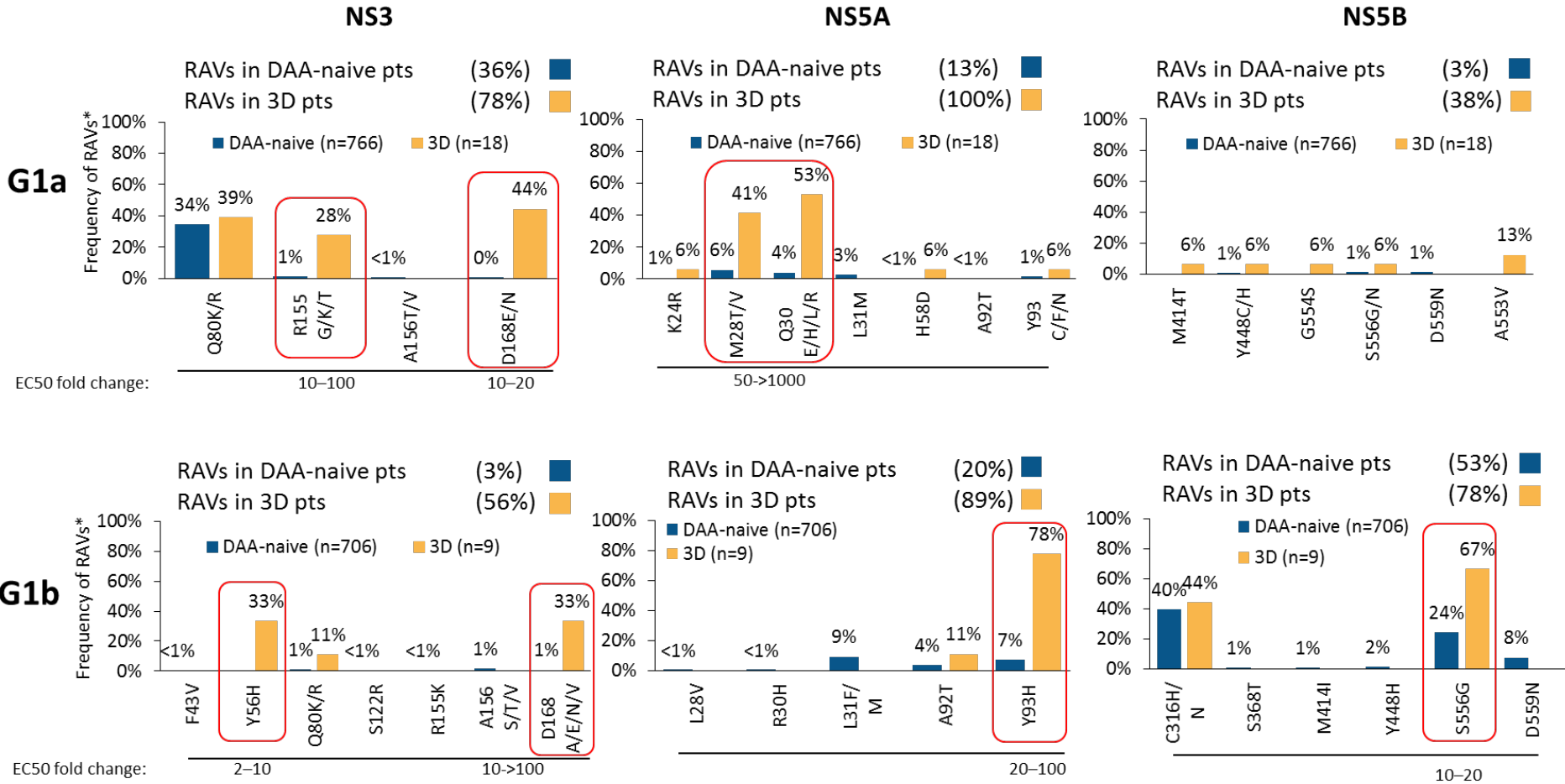


G4



Frequency and characteristics of RAVs in treatment-naïve and DAA-experienced patients - European RAVs database:

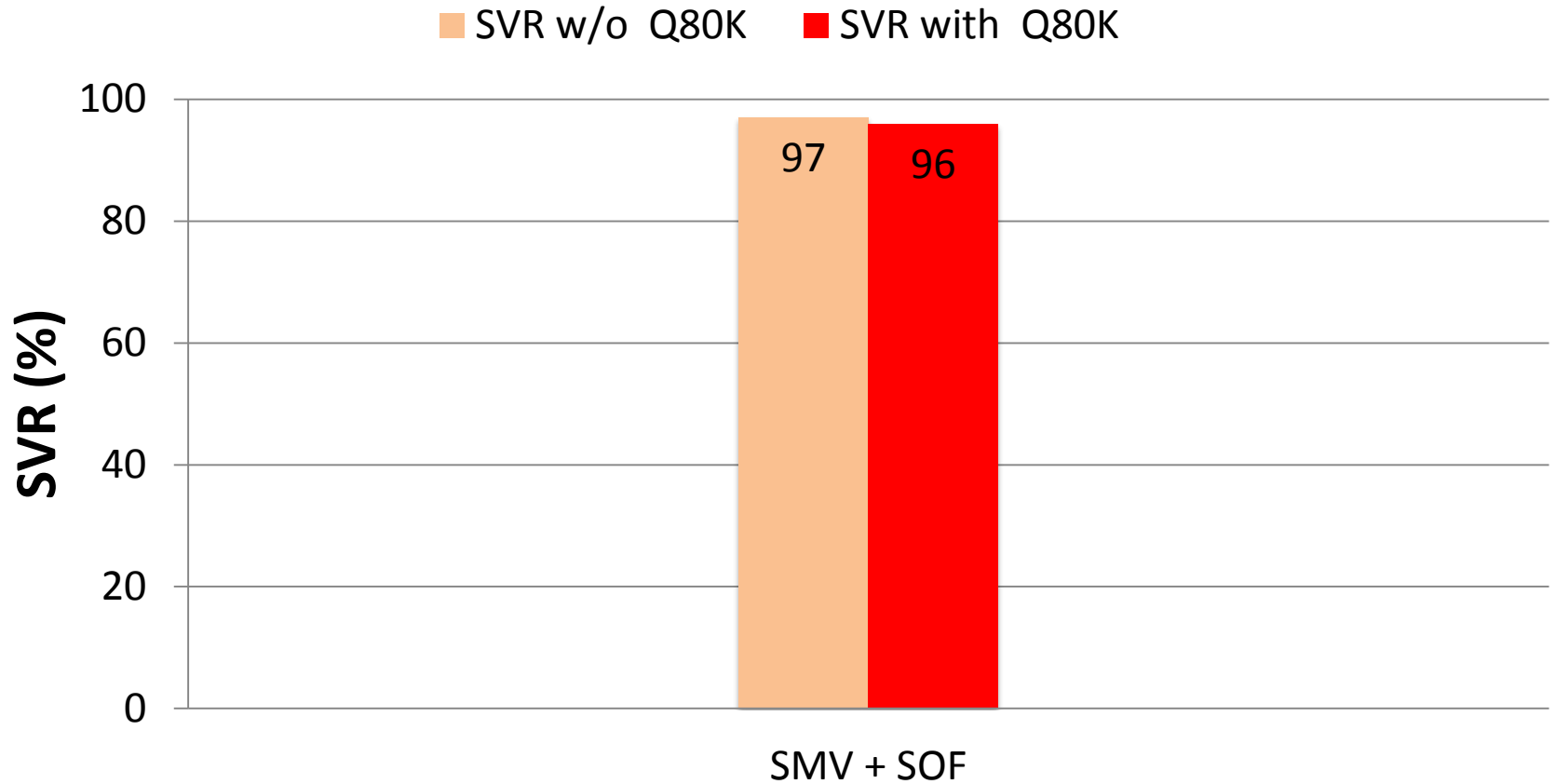
RAVs in 3D failures



How to define a RAV ?

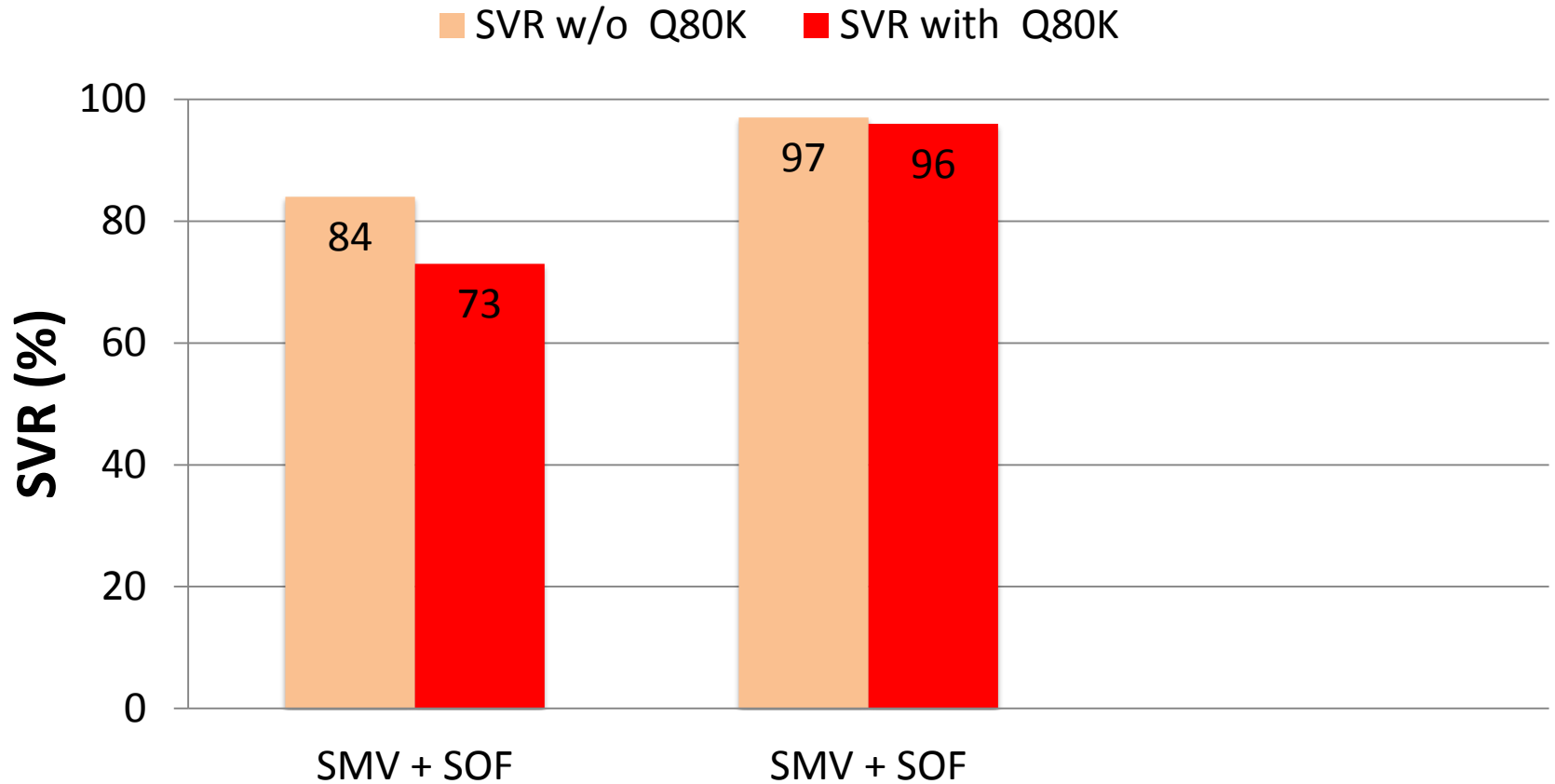
- RAV per specific drug or drug class
(e.g. resistance against ledipasvir or any NS5A inhibitor)
- Genotypic backbone *in vitro* and *in vivo*
(e.g. genotype 1a or 1b replicons, other genotypes)
- Level of reduced susceptibility *in vitro*
(e.g. any aa change vs >2.5-fold vs >100-fold)
- Methodology of RAV detection (sensitivity threshold)
(e.g. deep sequencing with a cut-off at 1% or populations sequencing with a cut-off at 15-25%)
- Impact of RAVs on SVR rates
(e.g. combination partner, treatment duration, stage of disease)

Simeprevir + Sofosbuvir in GT1a-infected patients (TN+TE): Impact of the NS3 RAV Q80K



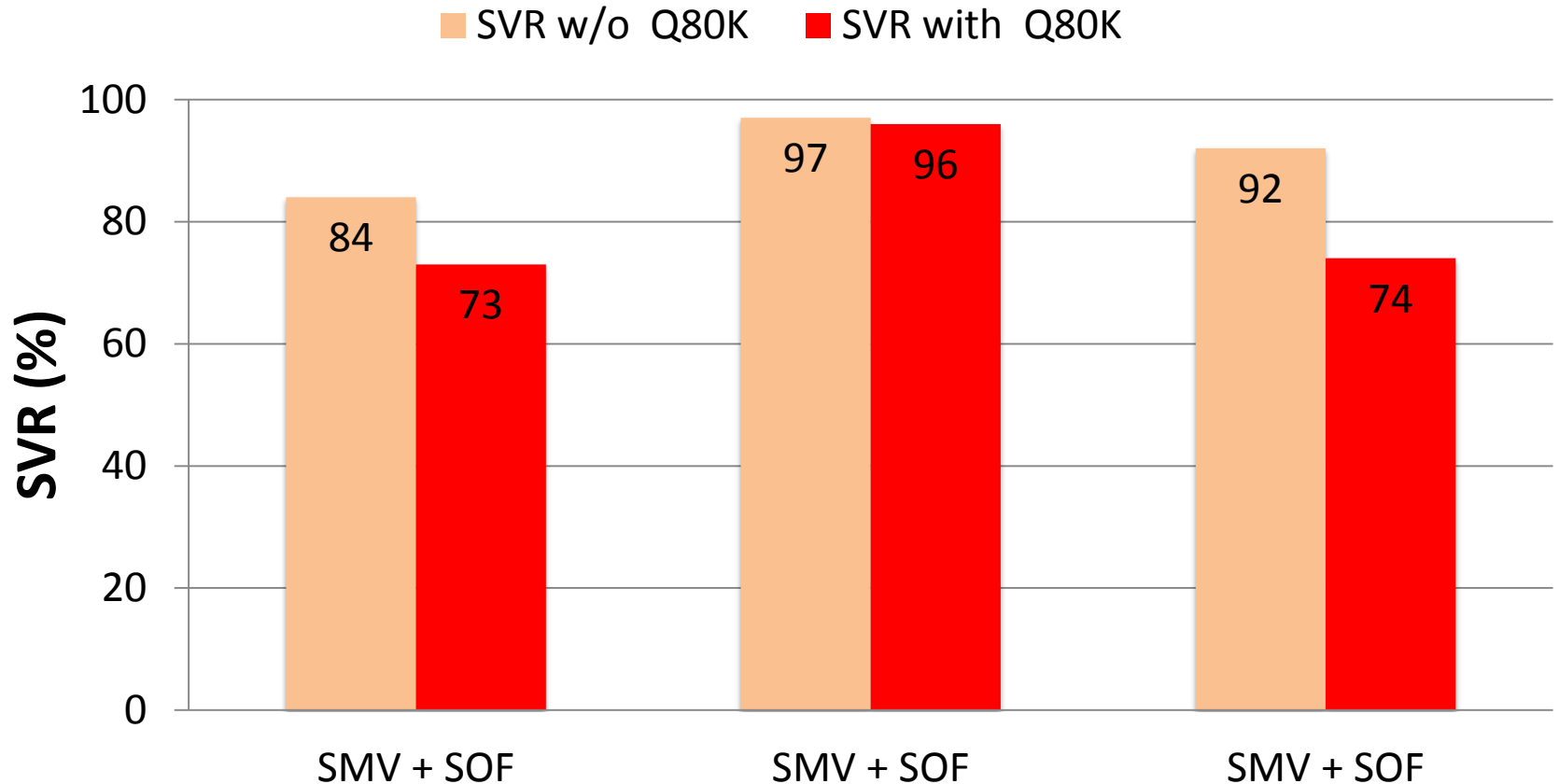
Tx duration		12 weeks	
Cirrhosis		No	
Q80K frequency		40%	

Simeprevir + Sofosbuvir in GT1a-infected patients (TN+TE): Impact of the NS3 RAV Q80K



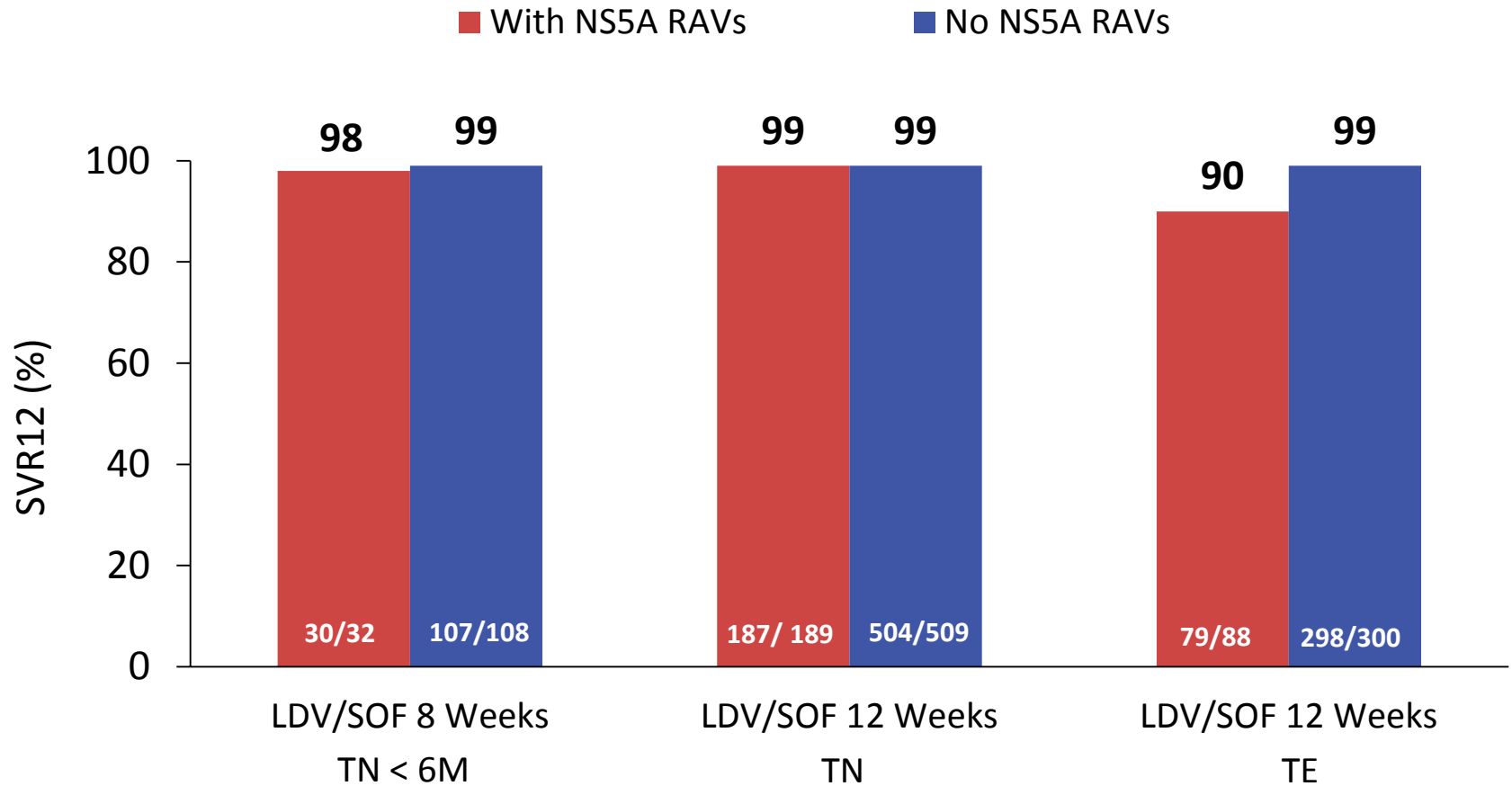
Tx duration	8 weeks	12 weeks	
Cirrhosis	No	No	
Q80K frequency	42%	40%	

Simeprevir + Sofosbuvir in GT1a-infected patients (TN+TE): Impact of the NS3 RAV Q80K



Tx duration	8 weeks	12 weeks	12 weeks
Cirrhosis	No	No	Yes
Q80K frequency	42%	40%	47%

SVR12 Rates by Treatment Regimen (LDV/SOF) and Duration: Patients without Cirrhosis



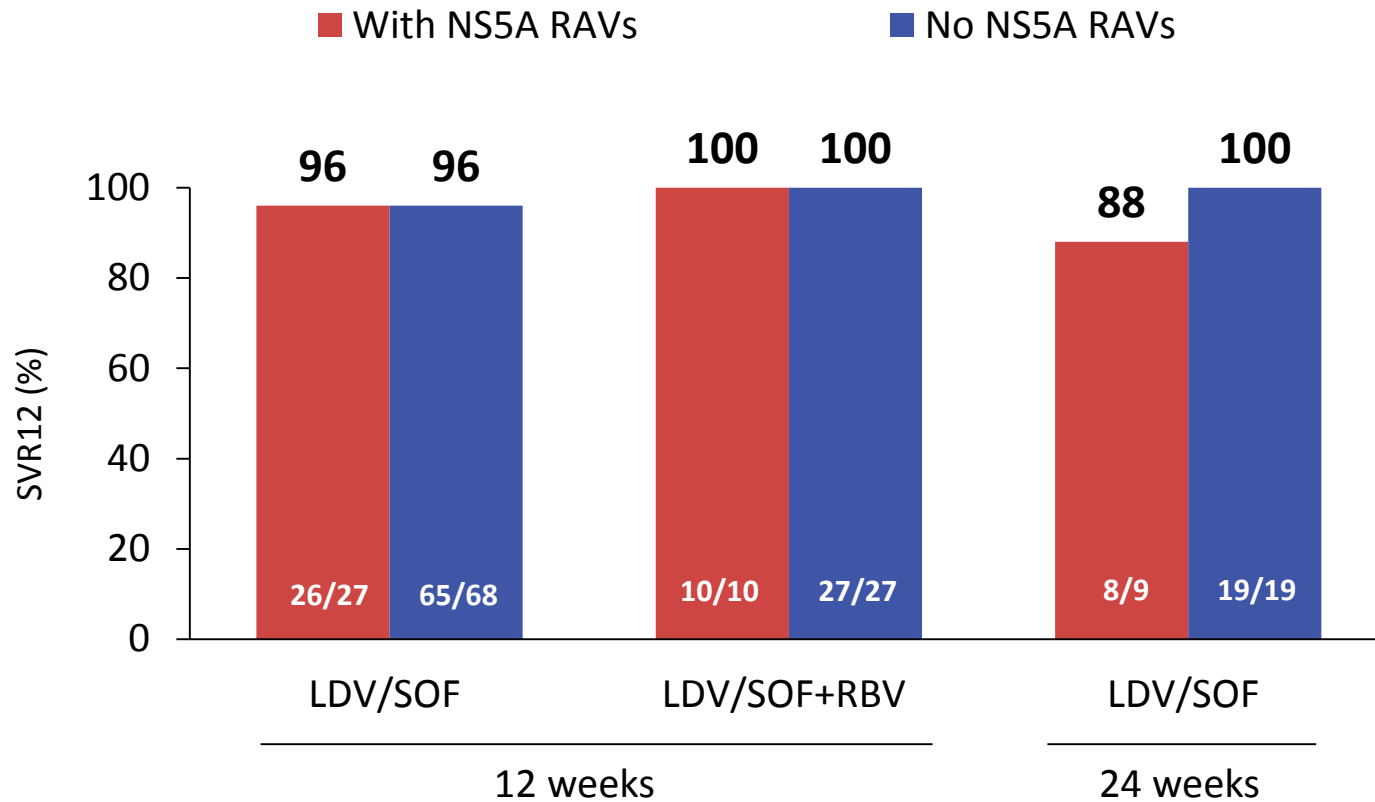
Studies included for analysis:

LDV/SOF 8 weeks: GS-US-337-0118 (LONESTAR 1), GS-US-337-0108 (ION-3); **LDV/SOF 12 Wks TN:** GS-US-GS-US-334-1274 (Bleeding Disorder), GS-US-337-0102 (ION-1), GS-US-337-0108 (ION-3), GS-US-337-0113 (Japan 1), GS-US-337-0115 (ION-4), GS-US-337-0122 (Electron 2), GS-US-337-0131 (China), GS-US-337-0118 (LONESTAR 1), GS-US-337-1406, GS-US-337-1468 (LEPTON); **LDV/SOF 12 Wks TE:** GS-US-337-0109 (ION-2), GS-US-337-0113 (Japan 1), GS-US-337-0115 (ION-4), GS-US-337-0124 (SOLAR-2), GS-US-334-1274 (Bleeding Disorder), GS-US-337-0118 (LONESTAR 1), GS-US-337-0131 (China), GS-US-337-1406, GS-US-337-1468 (LEPTON)

Sensitivity threshold at 1% (deep sequencing)

Zeuzem et al., AASLD 2015

SVR12 Rates by Treatment Regimen (LDV/SOF) and Duration: **TN** Patients with Cirrhosis



Studies included for analysis:

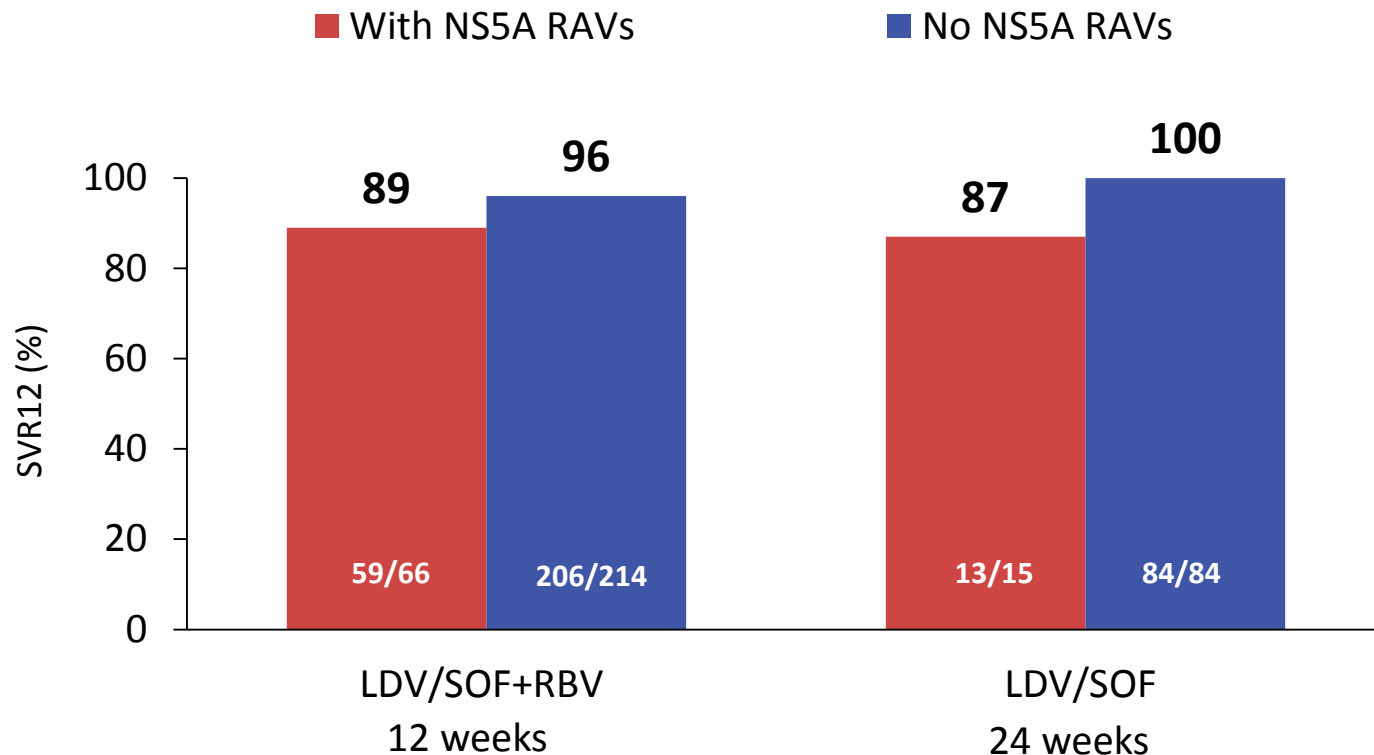
LDV/SOF 12 Wks: GS-US-334-1274 (Bleeding Disorder), GS-US-337-0102 (ION-1), GS-US-337-0113 (Japan 1), GS-US-337-0115 (ION-4), GS-US-337-0122 (ELECTRON-2), GS-US-337-0131(China), GS-US-337-1406;

LDV/SOF+RBV 12 Wks: GS-US-337-0102 (ION-1), GS-US-337-0113 (Japan 1), GS-US-337-0122 (ELECTRON-2); **LDV/SOF 24 Wks:** GS-US-337-0102 (ION-1), GS-US-334-1274 (Bleeding Disorder)

Sensitivity threshold at 1% (deep sequencing)

Zeuzem et al., AASLD 2015

SVR12 Rates by Treatment Regimen and Duration: **TE** Patients with Cirrhosis



Studies included for analysis:

LDV/SOF+RBV 12 Wks: GS-US-337-0109 (ION-2), GS-US-337-0113 (Japan 1), GS-US-337-0118 (LONESTAR-1), GS-US-337-0122 (ELECTRON-2), GS-US-337-0123 (SOLAR-1), GS-US-337-0124 (SOLAR-2), GS-US-337-1118 (Retreatment), P7977-0523 (ELECTRON); **LDV/SOF 24 Wks:** GS-US-337-0109 (ION-2), GS-US-337-0121 (SIRIUS), GS-US-334-1274 (Bleeding Disorder)

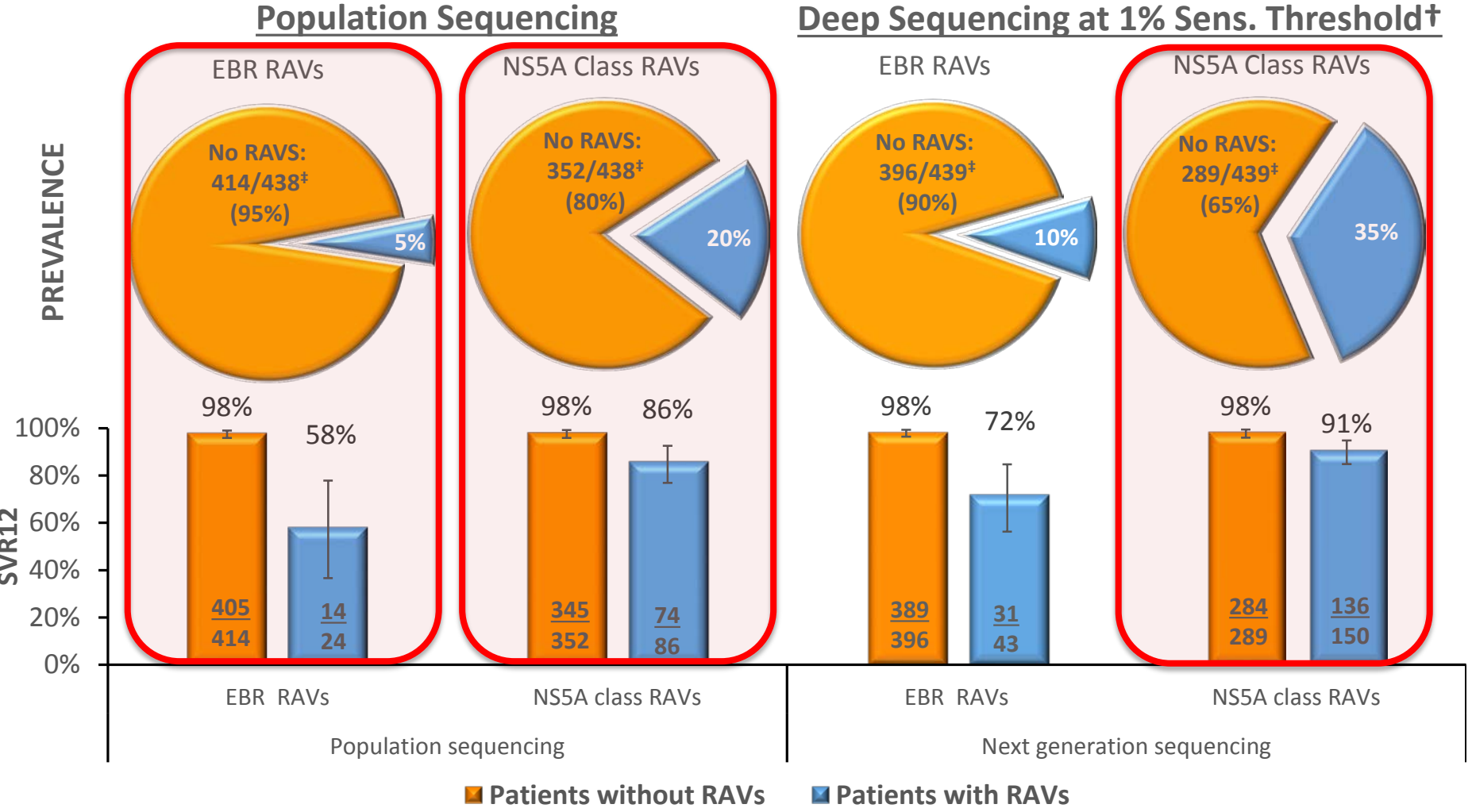
Sensitivity threshold at 1% (deep sequencing)

Zeuzem et al., AASLD 2015

How to define a RAV ?

- RAV per specific drug or drug class
(e.g. resistance against ledipasvir or any NS5A inhibitor)
- Genotypic backbone *in vitro* and *in vivo*
(e.g. genotype 1a or 1b replicons, other genotypes)
- Level of reduced susceptibility *in vitro*
(e.g. any aa change vs >2.5-fold vs >100-fold)
- Methodology of RAV detection (sensitivity threshold)
(e.g. deep sequencing with a cut-off at 1% or populations sequencing with a cut-off at 15-25%)
- Impact of RAVs on SVR rates
(e.g. combination partner, treatment duration, stage of disease)

Grazoprevir + Elbasvir for 12 wks in TN/relapse GT-1a patients: Impact of NS5A RAVs on SVR rates



[†]NGS with 1% ST supplemented by Population Sequencing when NGS not available. [‡] One GT1a was missing baseline population sequencing data but had baseline NGS data
 EBR RAV List = For GT1a: M/L28T/A/G, Q/R30E/H/R/G/K/L/D, L31M/V/F, H58D, or Y93C/H/N/S
 NS5A Class RAV List = Any variant from reference strain at NS5A position 24, 28, 30, 31, 32, 38, 58, 92 or 93

Effect of RAVs at Specific Baseline Positions on Likelihood to Achieve SVR₁₂

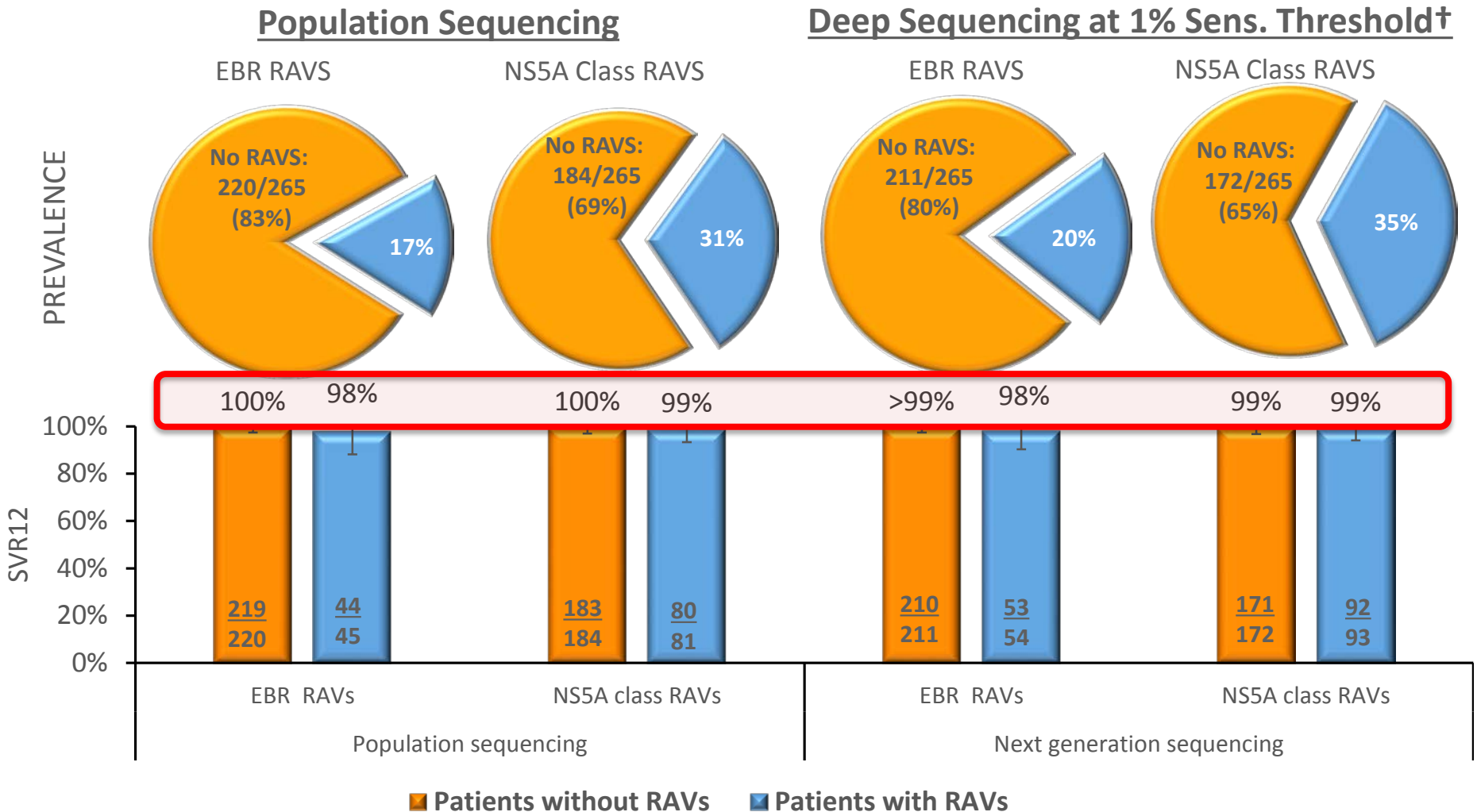
GT1a-Infected TN/TE Subjects given EBR/GZR 12 weeks (no RBV)

RAV Position	SVR12 Subjects with RAVs (1% ST NGS)	SVR12 Subjects With RAVs (Population Sequencing)
24	15/18 (83.3%)	4/4 (100.0%)
28	61/68 (89.7%)	29/33 (87.9%)
30	14/23 (60.9%)	4/10 (40.0%)
31	15/23 (65.2%)	5/13 (38.5%)
32	1/1 (100.0%)	--
38	9/9 (100.0%)	--
58	75/77 (97.4%)	48/49 (98.0%)
92	6/6 (100.0%)	3/3 (100.0%)
93	9/14 (64.3%)	5/8 (62.5%)

NGS using 1% ST supplemented by Population Sequencing when NGS not available

NS5A Class RAV List = Any variant from reference strain at NS5A position 24, 28, 30, 31, 32, 38, 58, 92 and 93

Grazoprevir + Elbasvir for 12 wks in TN/relapse GT-1b patients: Impact of NS5A RAVs on SVR rates

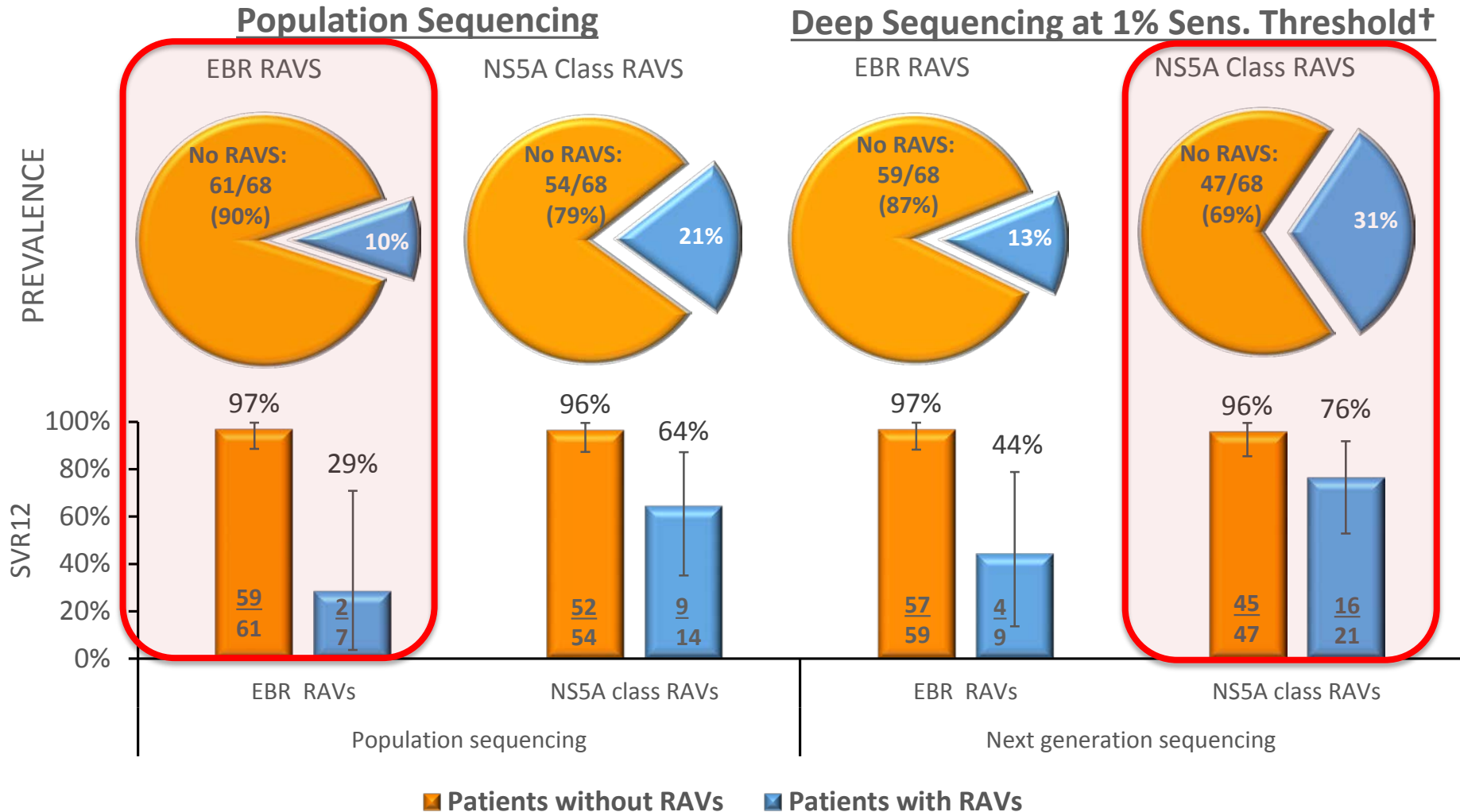


†NGS 1% ST supplemented by Population Sequencing when NGS not available

EBR RAV List = For GT1b: P32L, P58D or A92K or any variant at NS5A position 28, 30, 31 or 93

NS5A Class RAV List = Any variant from reference strain at NS5A position 24, 28, 30, 31, 32, 38, 58, 92 or 93

Grazoprevir + Elbasvir for 12 wks in non-responder GT-1a patients: Impact of NS5A RAVs on SVR rates

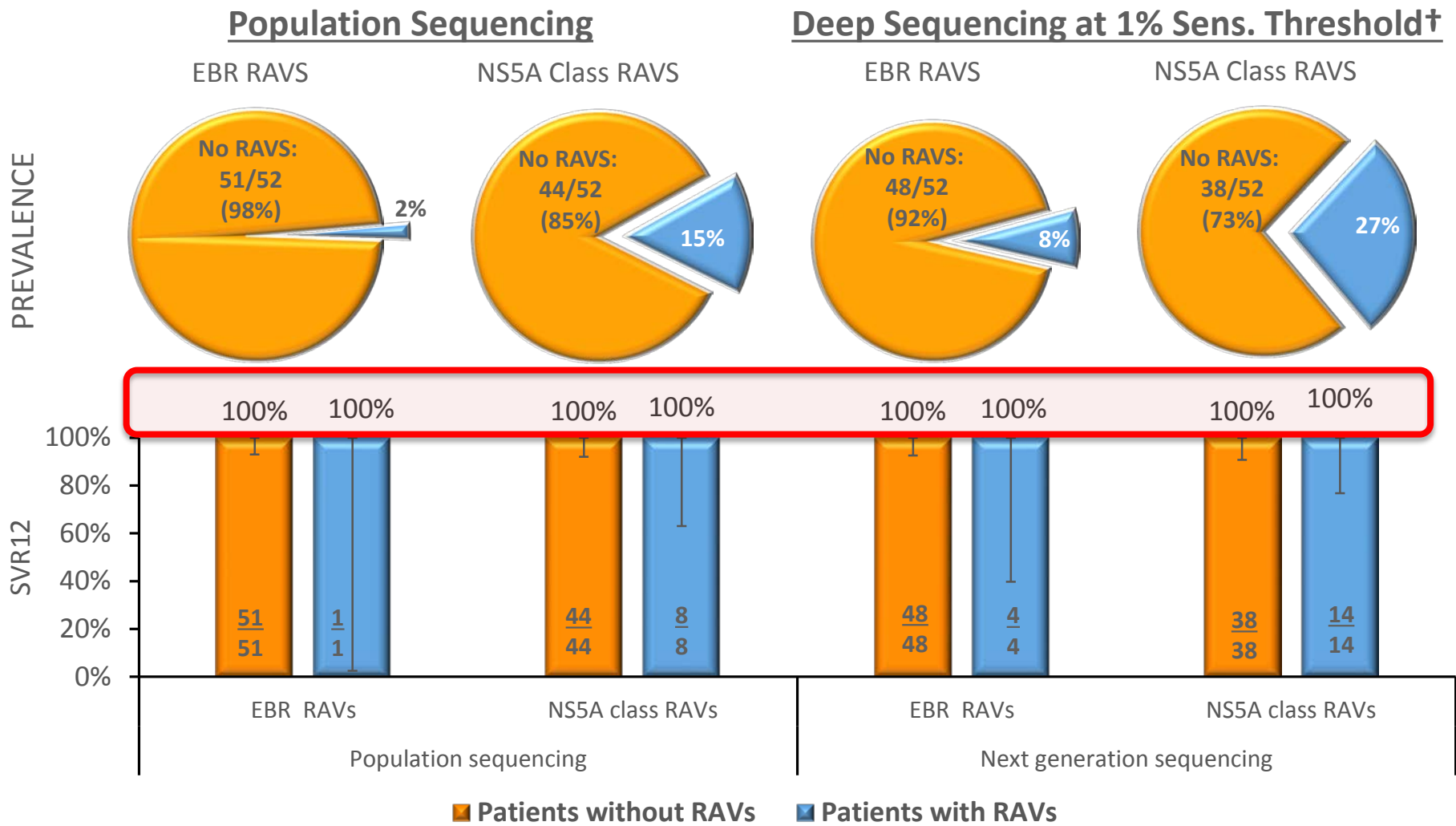


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Grazoprevir + Elbasvir + RBV for 16/18 wks in NR GT-1a patients: Impact of NS5A RAVs on SVR rates

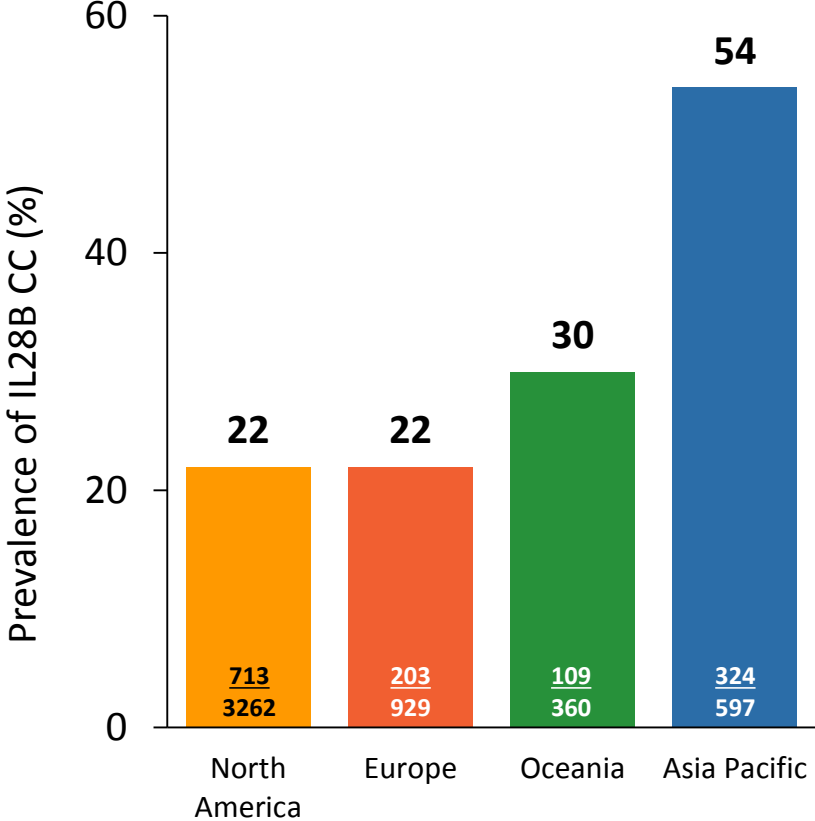
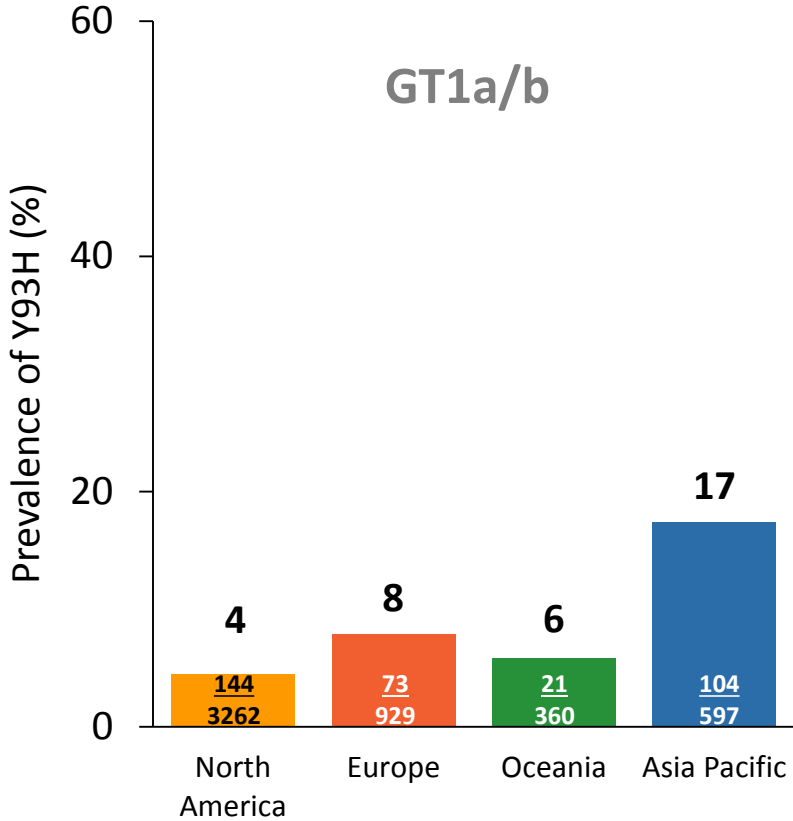


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 Jacobson et al., AASLD 2015

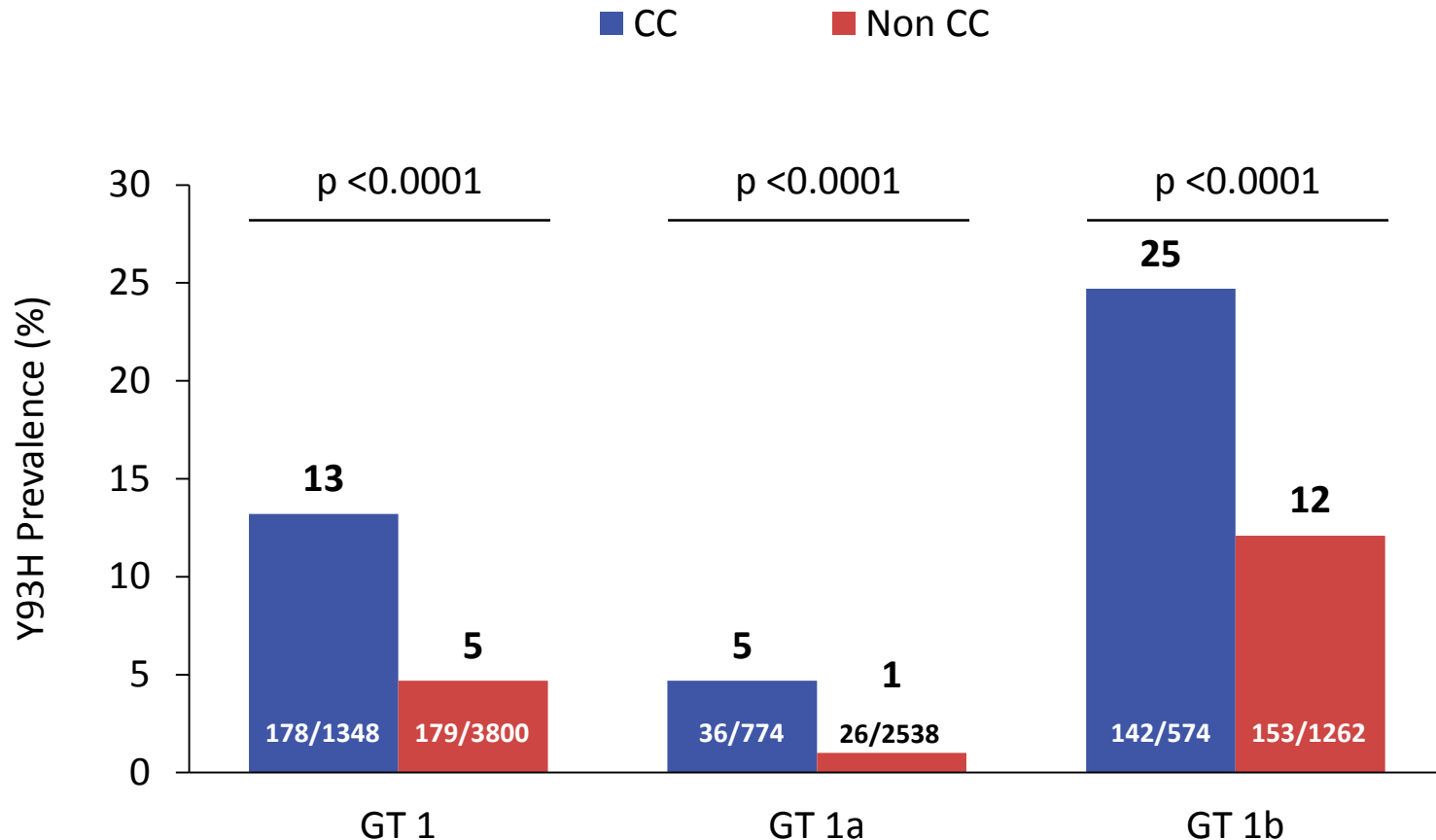
How to define a RAV best clinically?

- RAVs (single and combination) per specific drug
- Genotypic backbone *in vitro* and *in vivo*
- Level of reduced susceptibility *in vitro* >100-fold
- sensitivity threshold at 15-25% (population sequencing)
- Impact of RAVs on SVR rates in defined patient population and a defined regimen (DAA combination, treatment duration)
- and there is even more complexity

Y93H and IL28B CC prevalence by region

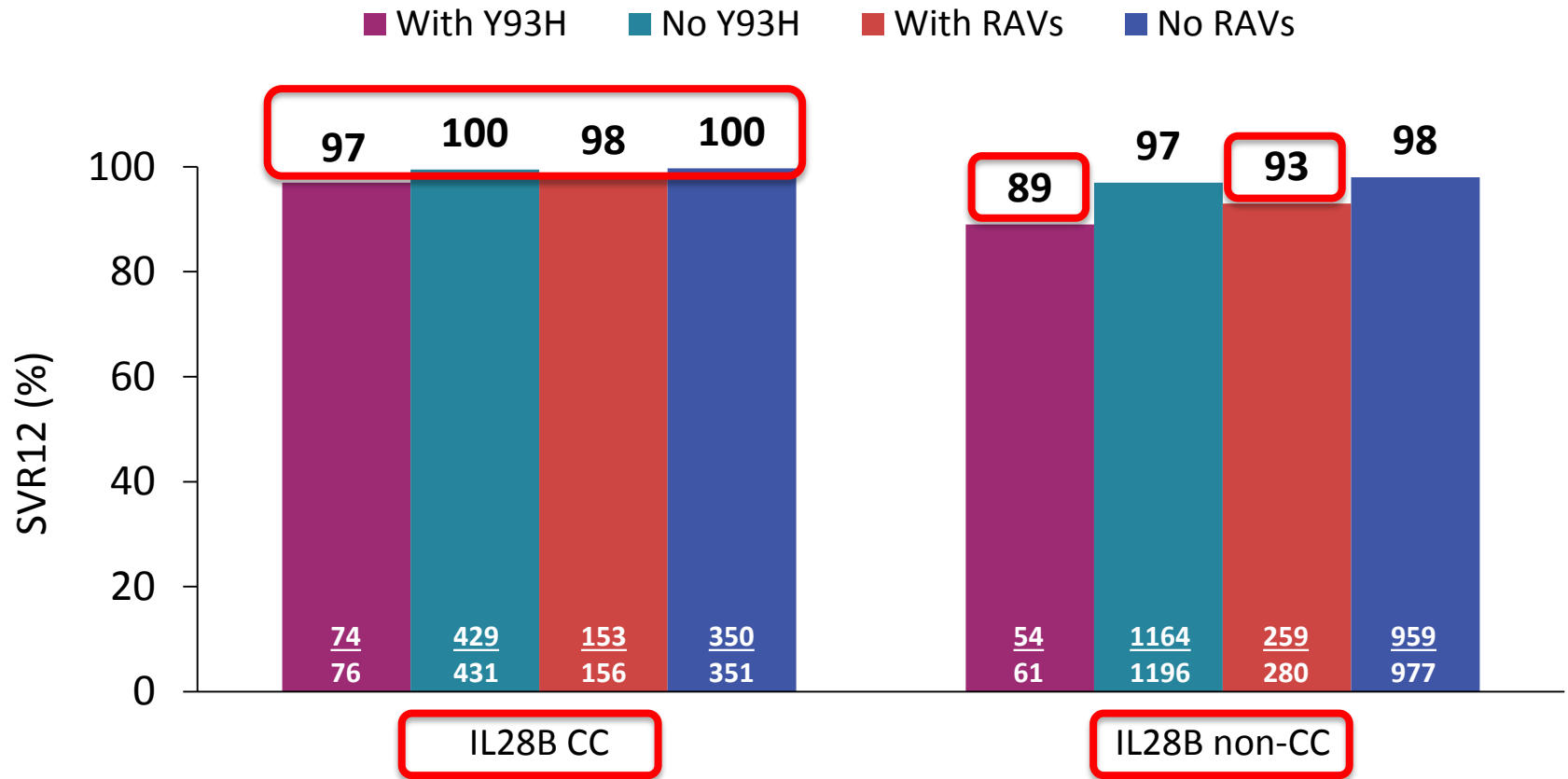


Association of Y93H and IL28B genotype



- ◆ 5,148 GT1 patients in Gilead HCV clinical trials with both NS5A sequencing and IL28B genotype data

SVR12 rates by Y93H or any NS5A RAV and IL28B genotype

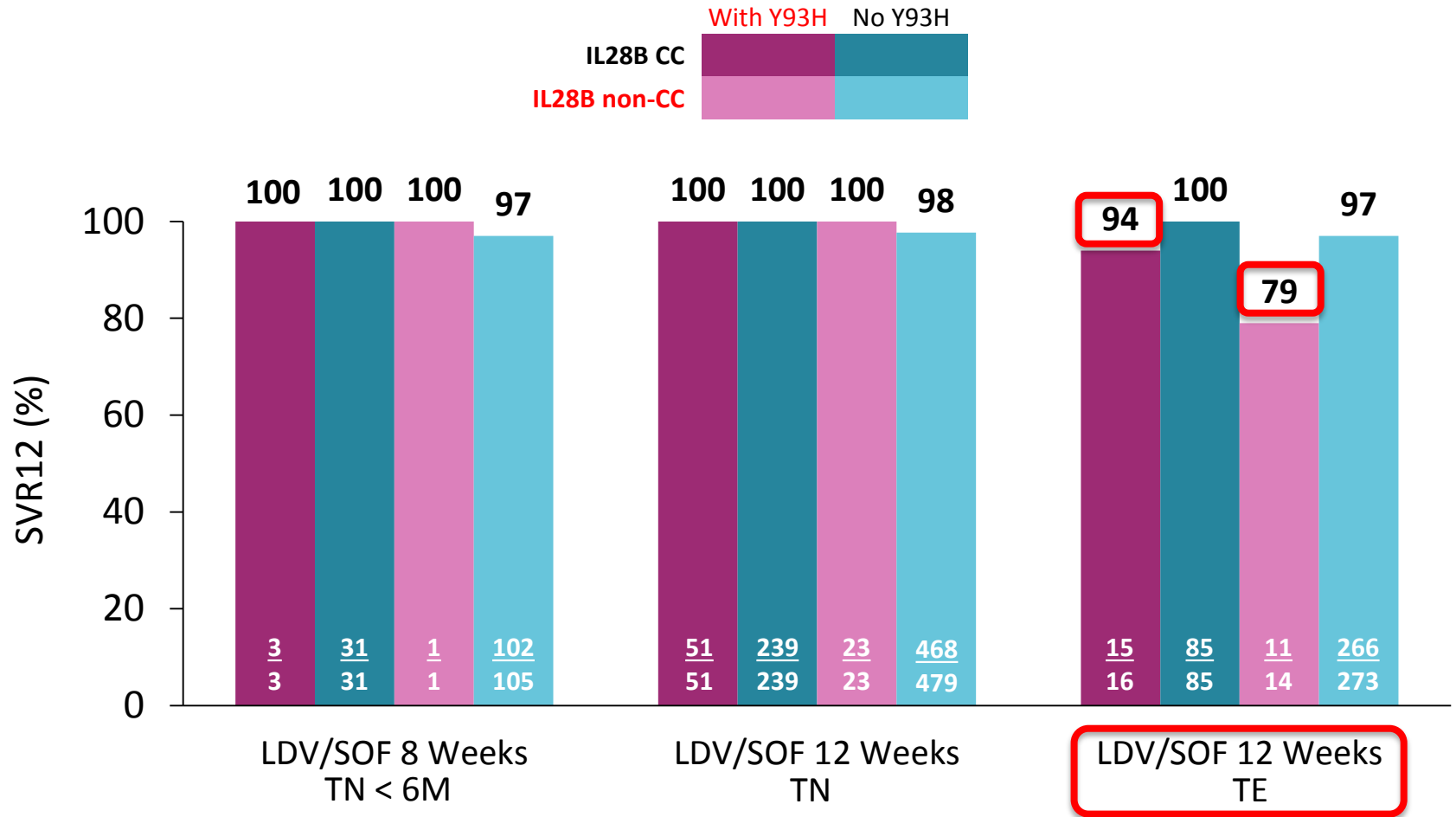


◆ SVR12 data pooled from all guideline recommended LDV/SOF regimens

Studies included for analysis:

GS-US-337-0118 (LONESTAR 1), GS-US-GS-US-334-1274 (Bleeding Disorder), GS-US-337-0102 (ION-1), GS-US-337-0108 (ION-3), GS-US-337-0113 (Japan 1), GS-US-337-0115 (ION-4), GS-US-337-0122 (Electron 2), GS-US-337-0131 (China), GS-US-337-1406, GS-US-337-1468 (LEPTON), GS-US-337-0109 (ION-2), GS-US-337-0124 (SOLAR-2), GS-US-334-1274 (Bleeding Disorder), GS-US-337-1406

SVR12 Rates by Treatment Regimen and Y93H/IL28B: Patients without Cirrhosis



Studies included for analysis:

GS-US-337-0118 (LONESTAR 1), GS-US-GS-US-334-1274 (Bleeding Disorder), GS-US-337-0102 (ION-1), GS-US-337-0108 (ION-3), GS-US-337-0113 (Japan 1), GS-US-337-0115 (ION-4), GS-US-337-0122 (Electron 2), GS-US-337-0131 (China), GS-US-337-1406, GS-US-337-1468 (LEPTON), GS-US-337-0109 (ION-2), GS-US-337-0124 (SOLAR-2), GS-US-334-1274 (Bleeding Disorder), GS-US-337-1406

It is useful to detect RAVs ?

- There is no “YES” or “No” answer
- First and most important: Methodology for detection of RAVs must **standardized** (and automated)
- Pharmaceutical industry must **fully publish available RAV data** in collaboration with academia
- Usefulness of RAV testing will be **patient population and treatment regimen** dependent
- RAV testing most likely not required in patient populations with SVR rates > 99%
- RAV testing most likely be clinical useful and cost-effective in population with **suboptimal SVR rates** (definition < 95% / < 90% ?) & if **population large enough**
 - Regimens w/o a very high barrier to resistance drug
 - Treatment-experienced patients (in particular when exposed to DAAs)
 - Patients with cirrhosis
 - When the shortest possible treatment duration is economically important
- **IL28B CC genotype** is significantly **associated with** a higher prevalence of **Y93H**. This association is functionally not yet understood
- The relevance of (in particular NS5A) RAVs, baseline HCV RNA levels, and IL28B genotype on treatment outcome requires further **intensive clinical research**

Treatment decisions in patients with RAVs

- Switch drug class
(exception: nucleosidic polymerase inhibitors)
- Treat longer (up to 24 weeks)
- Add ribavirin
- Use triple therapies (NI + PI + NS5A-Inhibitor)
- Second generation PIs and NS5A-Inhibitors
(e.g. Glecaprevir, Pibrentasvir)
- Peginterferon (??)

